Four Triterpene Oligoglycosides from the seeds of Aesculus Chinensis

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Abstract: A pair of novel geometrically isomeric triterpenoid saponins named escins IVa and IVb, together with known escins Ia and Ib, were isolated from the seeds of *Aesculus chinensis*. Their structures were determined on the basis of spectroscopic evidences and comparison.

Keywords: Aesculus chinensis, hippocastanaceae, escins Ia, Ib, IVa, Ivb, X-ray analysis.

The seeds of *Aesculus chinensis* Bge. (Hippocastanaceae) have been used as folk medicine for the treatment of distension and pain in chest and abdomen¹. Until now, there is no report on its chemical constituents. This paper describes structure elucidation of two new compounds, escins IVa **3** and IVb **4** on the basis of spectroscopic methods and comparison with escins Ia **1** and Ib 2^2 . Among them, Escin Ia **1** was studied by X-ray diffraction analysis.

Compound **3**, white powder, showed IR bands due to carboxyl and α , β -unsaturated ester at 1740,1713 and 1640 cm⁻¹ and strong absorption bands at 3409 and 1074cm⁻¹ suggestive of the oligoglycosidic structure². The positive-mode TOF-MS of **3** displayed quasimolecular ion peaks at m/z 1153 [M+Na]⁺ and 1130 [M]⁺ the same as **1** and **2**. ¹H and ¹³C NMR spectra of **3** were similar to those of **1** with presence of a tigloyl group, an acetyl group and a trisaccharide moiety. Carbon signals of C-21, C-22, C-17 and C-28, which differed from those of **1**, were assigned as in the **Table** with aid of HMQC and HMBC spectra. The occurrence of a tigloyl group at C-21 was confirmed by the cross peak between H-21 (δ 6.40) and the carbonyl carbon (δ 168.5) of the tigloyl group in HMBC spectrum. However no correlation signal was observed between the carbonyl carbon of the acetyl group at C-28 was deduced from significant shift for C-28, C-17, C-22 in ¹³C-NMR and C₂₈-H (δ 4.10, d, J=9.0Hz; 4.40, d, J=9.0Hz), C₂₁-H (δ 6.40, d, J=10.0Hz), C₂₂-H (δ 4.44, d, J=10.0Hz) in ¹H NMR compared with those (C₂₈-H: δ 3.35, d, J=10.5Hz; 3.58, d, J=10.5Hz. C₂₁-H: 6.56, d, J=10.0Hz; C₂₂-H: 6.24, d, J=10.0Hz) of **1**.

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Furthermore, the linkage site and sugar sequence were also identified by similar long-range correlations in HMBC spectrum just as in **1**: C-3 (90.9) and H-1'(4.72), C-2' (79.4) and H-1'' (5.42) and C-4' (82.6) and H-1''' (4.79). Therefore **3** was identified as 21β -tigloyl-28-acetylprotoaescigenin-3 β -O- [β -D-glucopyranosyl (1-2)][β -D-glucopy-ranosyl (1-4)] - β -D-glucopyranosiduronic acid, named escin IVa.

Compound **4**, white powder, showed IR bands ascribable to carboxyl and α , β -unsa- turated ester at 1740, 1709 and 1640 cm⁻¹ and strong absorption bands at 3414 and 1074 cm⁻¹ suggestive of the oligoglycosidic structure. The negative-mode TOF-MS of **4**.





showed quasimolecular ion peaks at m/z 1152 [M-H+Na]⁻ and 1130 [M]⁻. Comparison with ¹H and ¹³C NMR spectra of **3** and **4** led to the conclusion that an angeloyl group [¹H-NMR: δ 5.84 (1H, dq like, H-3^{'''}), 1.98 (3H, d, J=7.0Hz, Ang-4^{''''}) and 1.93 (3H, s, Ang -5^{''''}); ¹³C-NMR: as shown in the **Table**] presented in place of a tigloyl group in the structure. Similar correlation relationships were also observed in HMBC and ¹H-¹H NOESY spectra. Therefore, **4** was elucidated as 21β-angeloyl-28-acetylprotoaescigenin-3β-O- [β-D-glucopyranosyl (1-2)] [β-D-glucopyranosyl (1-4)]-β-D-glucopyranosi-duronic acid , named escin IVb, a geometric isomer of **3**.

The existance of escins IVa and IVb in *Aesculus hippocastanum* has been foreseen by Griffini *et al*³ deduced from combined LC-MS. But it was for the first time that they were fully characterized by spectroscopic methods.

С	Ia	Ib	IVa	IVb	С	Ia	Ib	IVa	IVb
1	38.3	38.1	38.3	38.3	29	29.4	29.2	29.4	29.6
2	26.4	26.2	26.3	26.3	30	20.0	20.0	20.0	20.1
3	90.9	90.8	90.9	91.0	1'	104.5	104.3	104.3	104.5
4	43.5	43.3	43.4	43.5	2'	79.5	78.6	79.4	79.7
5	55.9	55.8	55.9	55.9	3'	76.3	76.3	76.4	76.5
6	18.4	18.2	18.3	18.4	4'	81.7	81.9	82.6	82.6
7	33.1	32.9	33.0	33.0	5'	75.6	75.3	75.5	75.5
8	39.8	39.8	39.7	39.7	6'	170.9	170.9	170.6	170.6
9	46.6	46.4	46.5	46.6	1"	104.1	103.8	103.9	104.1
10	36.4	36.0	36.3	36.2	2"	75.6	75.3	75.5	75.5
11	23.9	23.7	23.9	23.9	3"	78.4	78.1	78.0	78.2
12	123.0	123.1	123.2	123.2	4"	69.6	69.4	69.5	69.6
13	142.7	142.5	142.6	142.5	5"	78.0	77.7	78.0	78.1
14	41.5	41.3	41.6	41.6	6"	61.4	61.3	61.4	61.4
15	34.5	34.3	34.4	34.5	1'''	104.5	104.3	104.3	104.2
16	67.9	67.7	67.4	67.5	2'''	74.8	74.6	74.8	74.7
17	47.8	47.7	46.9	46.8	3'''	78.2	77.9	78.0	78.2
18	39.9	39.6	40.4	40.4	4'''	71.4	71.1	71.2	71.2
19	47.1	46.9	46.9	46.9	5'''	77.9	77.5	77.8	77.8
20	36.2	36.0	36.2	36.0	6'''	62.2	61.9	62.0	61.9
21	79.3	79.3	81.4	81.0	1""	167.9	167.6	168.5	168.4
22	74.1	74.6	71.0	70.9	2""	129.3	128.6	129.6	129.3
23	22.3	22.2	22.3	22.3	3""	136.8	137.0	136.2	136.1
24	63.2	63.0	63.1	63.1	4""	14.1	15.3	14.0	15.8
25	15.4	15.7	15.4	15.4	5""	12.3	20.8	12.3	20.9
26	16.6	16.4	16.7	16.7	1'''''	170.9	170.9	170.6	170.6
27	27.3	27.1	27.2	27.3	2"""	20.8	20.8	20.6	20.7
28	63.6	63.5	66.3	66.3					

Table. ¹³C-NMR Data of Escins Ia, Ib, IVa and Ivb

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